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# Denitrification in a Shallow Aquifer Underlying a Dairy Farm: New Approaches to Characterization and Modeling: UCRL-PRES-207404



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# We must understand denitrification to simulate nitrate transport

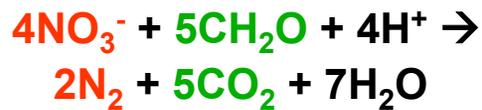


## ■ Denitrification requires

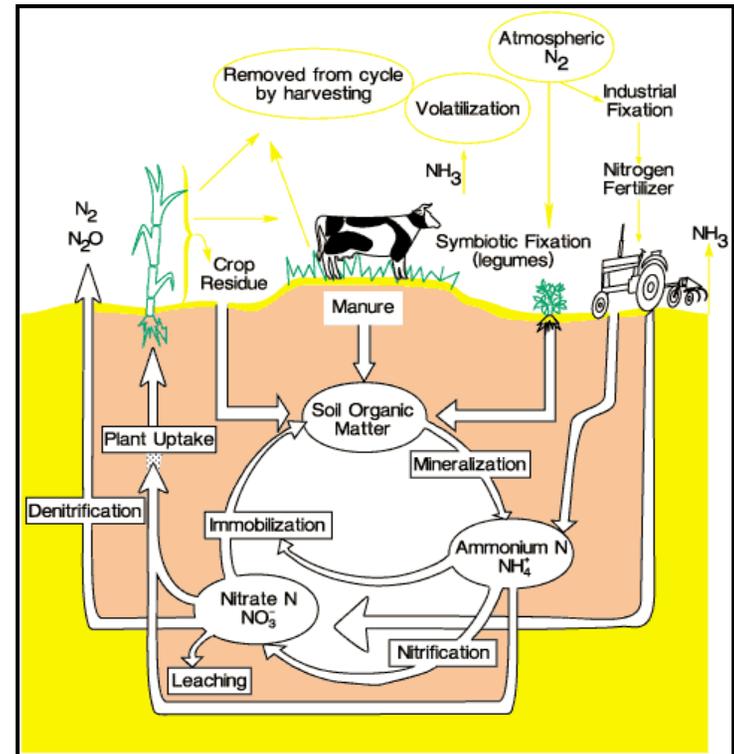
- Denitrifying bacteria
- Low oxygen conditions (< 0.6 mg/L)
- An electron donor

$\text{NO}_3^- \rightarrow$	$\text{NO}_2^- \rightarrow$	$\text{NO} \rightarrow$	$\text{N}_2\text{O} \rightarrow$	$\text{N}_2$
Nitrate (+5)	Nitrite (+3)	Nitric Oxide (+2)	Nitrous Oxide (+1)	Nitrogen (0)

## ■ Heterotrophic denitrification



## ■ Autotrophic denitrification



# The Goal is to predict the future concentration of nitrate at a receptor (drinking water well)



**Rate of nitrate change** at a given receptor = **Dispersion:** mechanical dispersion, molecular diffusion + **Advection:** bulk groundwater flow + **Source/sinks:** mass loading, reactions

$$\frac{\partial c_{NO_3^-}}{\partial t} = \frac{\partial \left( D \frac{\partial c_{NO_3^-}}{\partial x} \right)}{\partial x} - \frac{\partial (vc_{NO_3^-})}{\partial x} + \sum_i S_{NO_3^-} + R_{NO_3^-}$$

**Local groundwater velocity:**  
(Darcy's Law; age gradient)

**Source loading:**  
nitrate migration to the water table

**Denitrification rate expression:**  
microbial kinetics

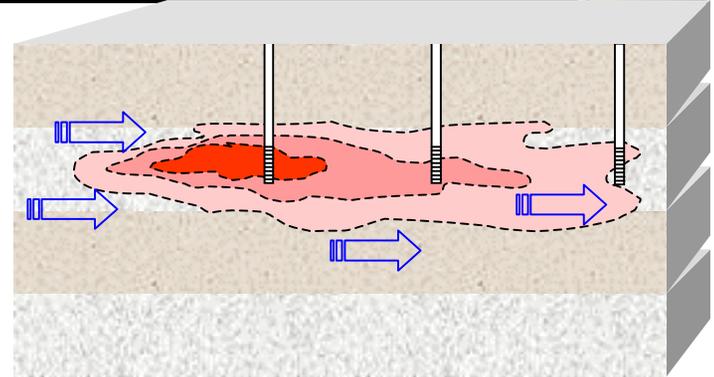
Parameters that may vary in space and hence will be modeled stochastically

# Quantifying denitrification requires a multi-disciplinary approach



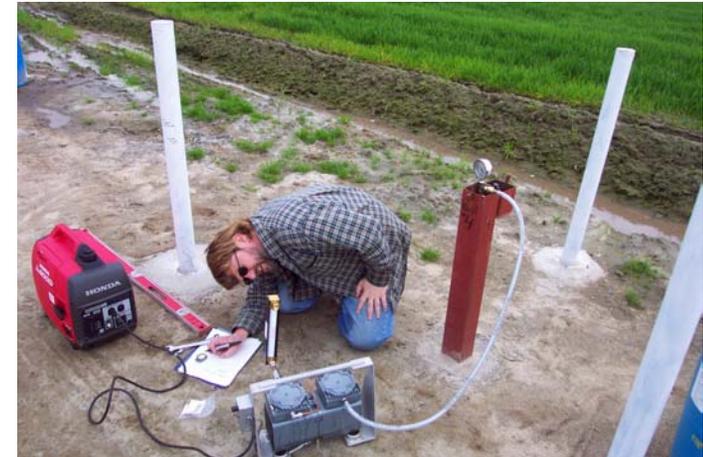
## ■ Characterization of groundwater flow

- Historical and current WLs, pump tests
- Tritium-helium age dating
- Stable isotopes of the water molecule
- Vadose zone instrumentation



## ■ Characterization of nitrate biogeochemistry & source

- Real-time quantitative PCR
- Microbial kinetics
- **Excess nitrogen**
- Stable isotopes of nitrate
  - $\delta^{15}\text{N}$  and  $\delta^{18}\text{O}$  of  $\text{NO}_3$
- Co-contaminants as source tracers



## ■ Modeling groundwater flow and chemistry

- Stochastic models (for spatial heterogeneity)
- Streamline & gridded flow & transport models
- Reactive transport

# We are using a molecular biology approach to measuring denitrification rates in the field



**Denitrifier population**

(# cells/volume from qPCR analysis of field sample)

X

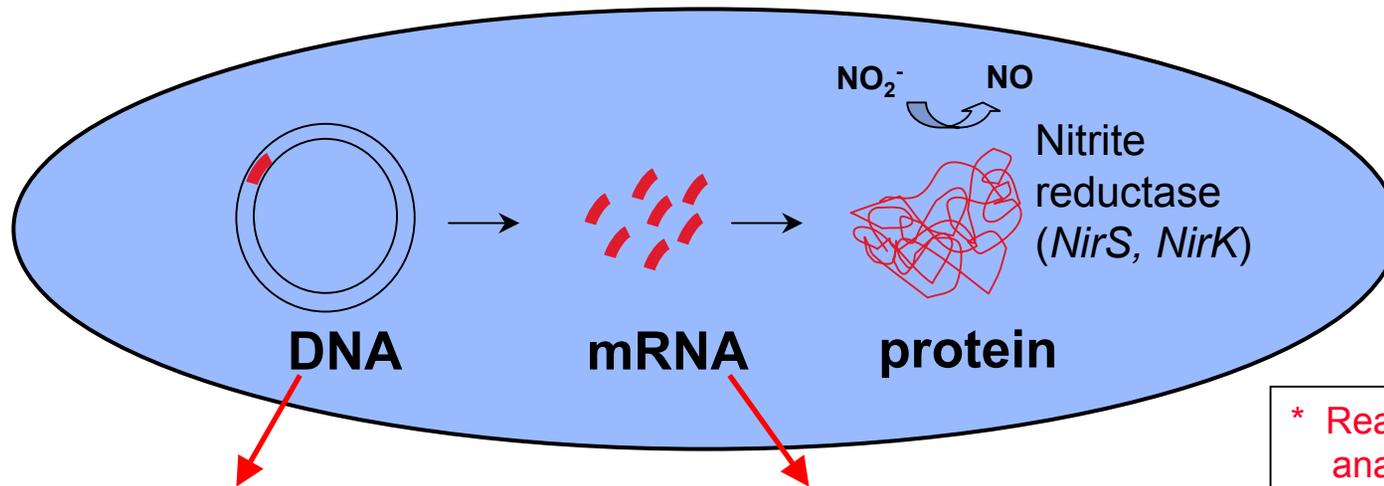
**Population-normalized denitrification rate**

( $\mu\text{mol nitrate}/\text{time}/\text{cell}$ , determined in the laboratory)

=

**Aquifer denitrification rate**

( $\mu\text{M nitrate}/\text{time}$ , input for transport model)



Population (# gene copies)\* x  
Specific rate (rate/cell)\*\* =  
Potential Denitrification Rate

# mRNA copies\* x  
Rate/mRNA copy\*\* =  
Actual denitrification rate

\* Real-time PCR analysis of aquifer sample  
\*\* Lab-determined denitrification rate

- Quantitative real-time PCR is a rapid, sensitive, and highly specific method that can be used to quantify denitrifying bacterial populations based on a diagnostic, **functional** gene.

# Key Aspects of Method Development: An assay for denitrifier population & population-normalized denitrification rate constants



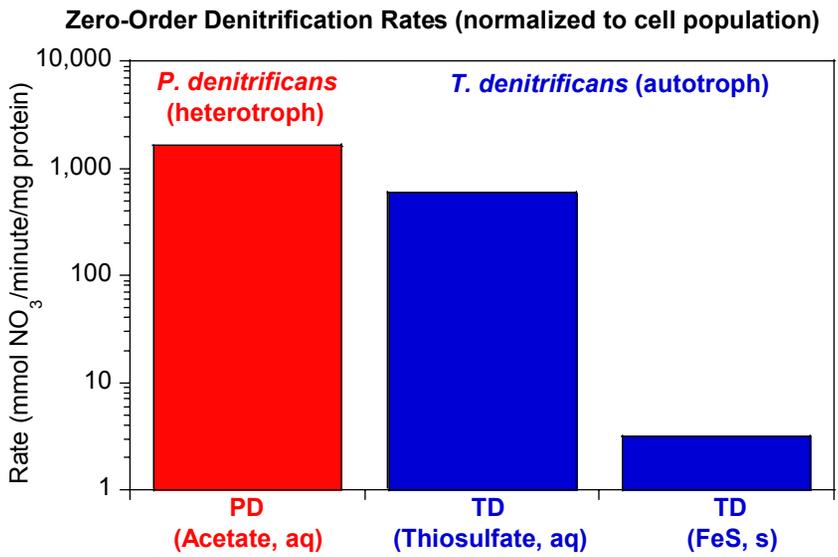
## Quantitative real-time PCR: Determining denitrifier populations

- Determined sequence for an **autotrophic** denitrification gene (*nirS*).
- High homology to **heterotrophic** *nirS* genes.
- We are now designing a **functional** test for **denitrifier cell populations** using qPCR.

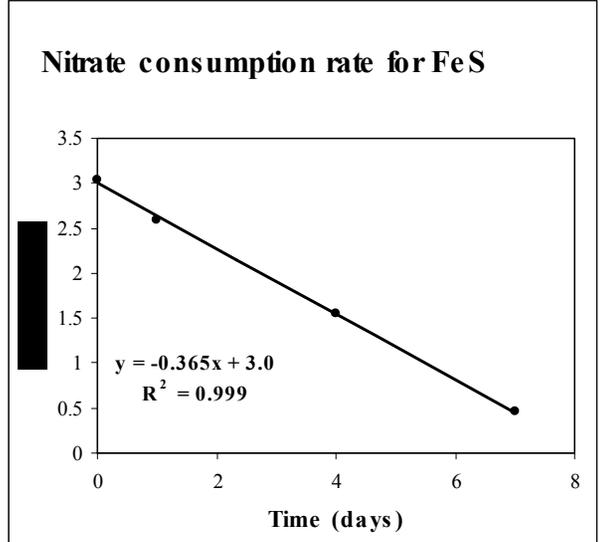
	(1252)	1252	1260	1270	1280	1290	1300
<b>Thiobacillus denitrificans</b>	(4)	TTCCCTGCATGAGGGCGGCTGGGA	TTCCAGGCACCGGTACTTTCATGTCGGGGCCAAACG				
Burkholderia cepacia	(1067)	TTCCCTCCACGACGGCGGCTGGGACAGCAGCCACCGGTACTTTCATGACCGGGCCAAACG					
Pseudomonas aeruginosa	(1048)	TTCCCTCCACGACGGCGGCTGGGACAGCAGCCACCGGTACTTTCATGACCGGGCCAAACG					
Pseudomonas sp. BH11.6	(188)	TTCCCTGCATGAGGGCGGCTGGGACAGCAGCCACCGGTACTTTCATGACGGGGCCAAACG					
Azarcus toluyliticus	(243)	TTCCCTGCATGAGGGCGGCTGGGACAGCAGCCACCGGTACTTTCATGACGGGGCCAAACG					
Paracoccus pantotrophus	(1114)	TTCCCTGCACGACGGCGGCTGGGACAGCAGCCACCGGTACTTTCATGACCGGGCCAAACG					
Paracoccus denitrificans	(1252)	TTCCCTGCACGACGGCGGCTGGGACAGCAGCCACCGGTACTTTCATGACCGGGCCAAACG					
Roseobacter denitrificans	(203)	TTCCCTGCATGATGGCGGTCTGGACAGCAGCCACCGGTACTTTCATGACGGGGCCAAACG					
Alcaligenes eutrophus	(1227)	TTCCCTGCATGAGGGCGGCTGGGACAGCAGCCACCGGTACTTTCATGACGGGGCCAAACG					
Azospirillum brasiliense	(200)	TTCCCTGCACGACGGCGGCTGGGACAGCAGCCACCGGTACTTTCATGACGGGGCCAAACG					
Alcaligenes faecalis	(200)	TTCCCTGCACGACGGCGGCTGGGACAGCAGCCACCGGTACTTTCATGACGGGGCCAAACG					
Pseudomonas stutzeri	(1003)	TTCCCTGCATGAGGGCGGCTGGGACAGCAGCCACCGGTACTTTCATGACGGGGCCAAACG					
Thauera aromatica	(240)	TTCCCTCCATGATGGCGGTGGGA	TGCGTCGGGTGTTACTTTCATGTTGGGGCCAAACG				
Acidovorax sp.2FB7	(240)	TTCCCTCCATGACGGTGGCTGGGA	TGCATCGGGCCGCTACTTTCATGTTGGGGCCAAACG				
Flavobacterium sp.BH12.12	(13)	TTCCCTGCACGATGGTGGCTTGGACGGCTCAGGTTACTTTCATGTTGGGGCCAAACG					
<b>Consensus</b>	(1252)	TTCCCTGCATGAGGGCGGCTGGGAC	CTCC	ACCGCTACTTTCATG	CCGGGGCCAAACG		

Gene sequences for denitrification gene

## Microbial kinetics: Biomass-normalized (“specific”) denitrification rate



The physical state of the electron acceptor (aqueous or solid) is major control on rate.

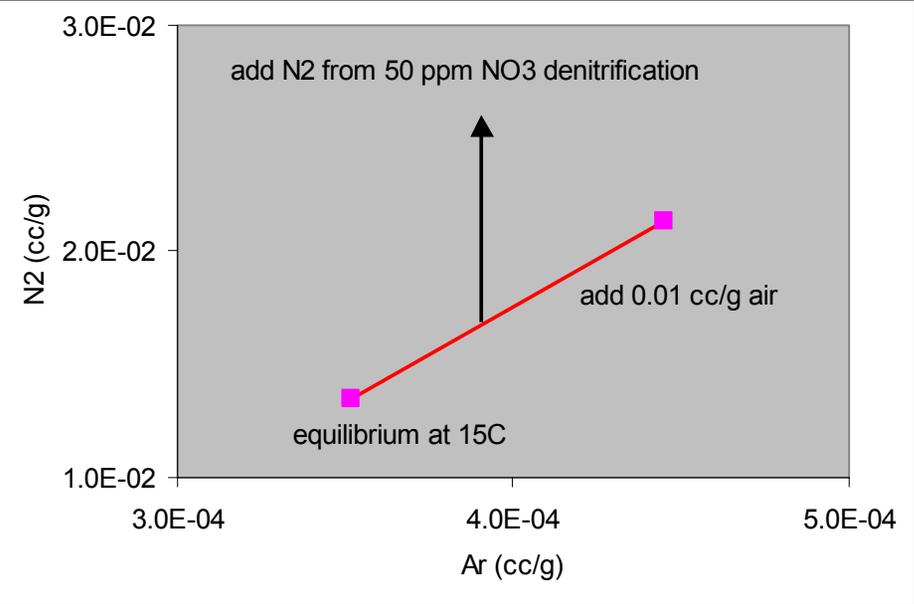


Denitrification rates are zero-order with respect to nitrate concentration.



# Excess nitrogen measurements distinguish denitrification from dilution

$\text{NO}_3^- \rightarrow$	$\text{NO}_2^- \rightarrow$	$\text{NO} \rightarrow$	$\text{N}_2\text{O} \rightarrow$	$\text{N}_2$
Nitrate (+5)	Nitrite (+3)	Nitric Oxide (+2)	Nitrous Oxide (+1)	Nitrogen (0)



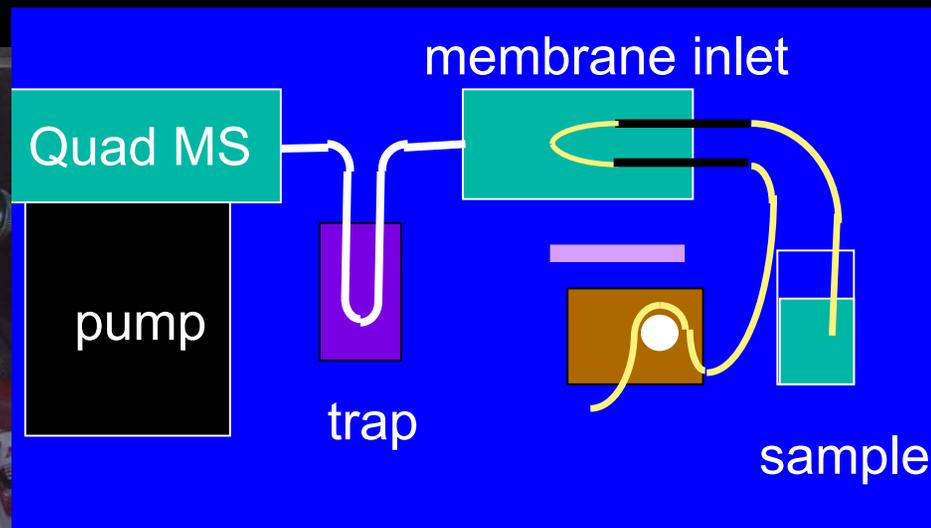
■ **The end product of denitrification is molecular nitrogen (N<sub>2</sub>, g)**

- Groundwater contains air above equilibrium solubility levels
- “Excess nitrogen” is the non-atmospheric N<sub>2</sub> component due to denitrification
- The atmospheric component is determined from the dissolved Ar concentration

■ **Excess N<sub>2</sub> allows quantification nitrate transformation**

- $F = 1 - \text{residual}/\text{initial nitrate}$   
Initial = residual + excess N<sub>2</sub>
- With age information, a bulk denitrification rate can be determined

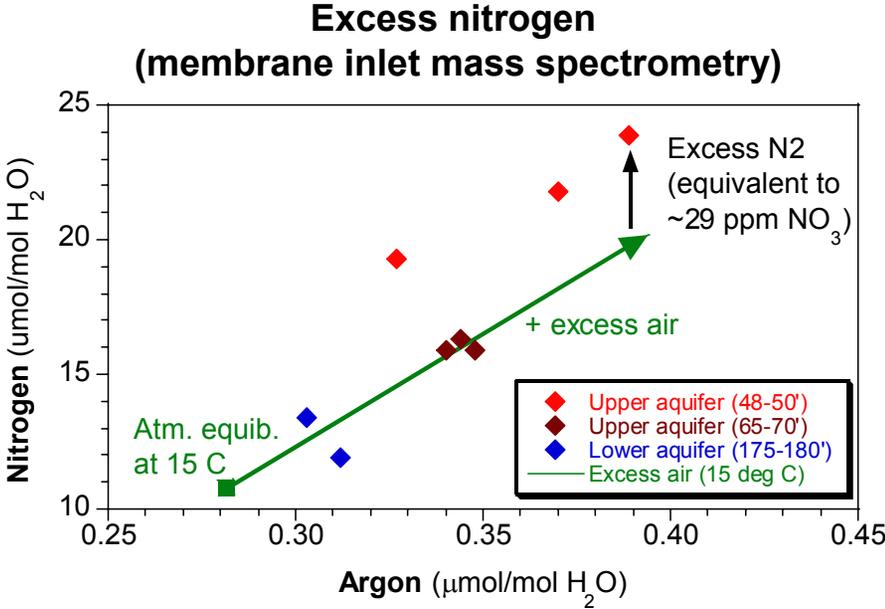
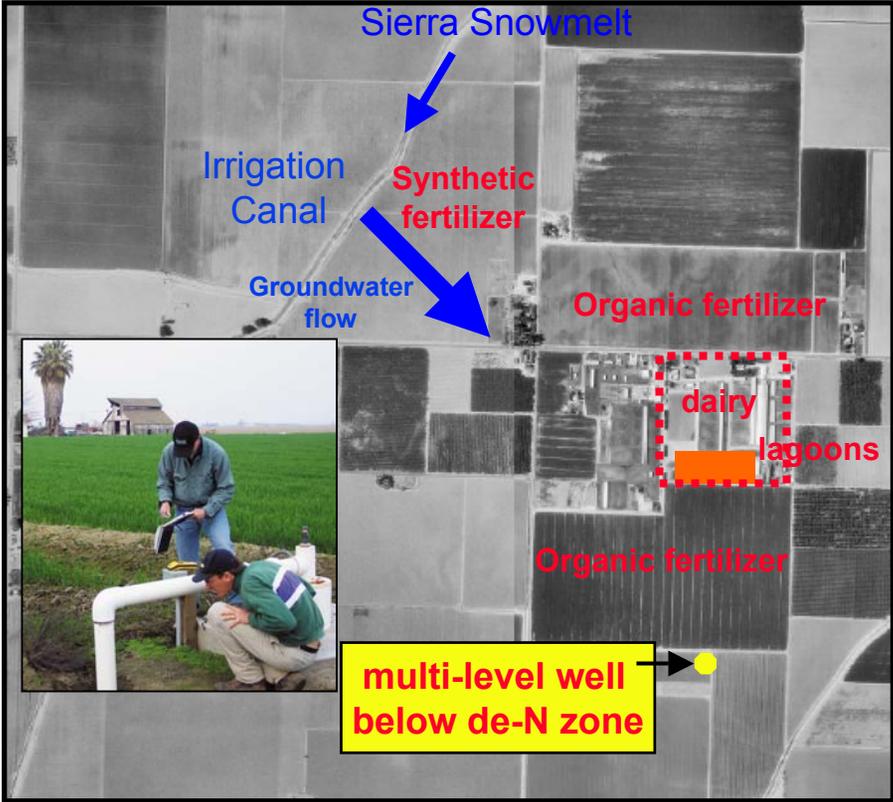
# The excess nitrogen approach



- **Membrane inlet mass spectrometry**
  - Measures nitrogen, argon, oxygen, carbon dioxide, and methane
  - Allows determination of excess nitrogen
  - Fast, portable and inexpensive



# Early results indicated that denitrification was taking place





# Several constituents were analyzed in the field during direct-push sampling

CPT-1

## Field methods

- Nitrate “sticks”
- Horiba water quality meter
  - DO, Conductivity, Temp, pH, ORP
- MIMS: Excess N<sub>2</sub>, CO<sub>2</sub>, CH<sub>4</sub>, excess air

## Samples

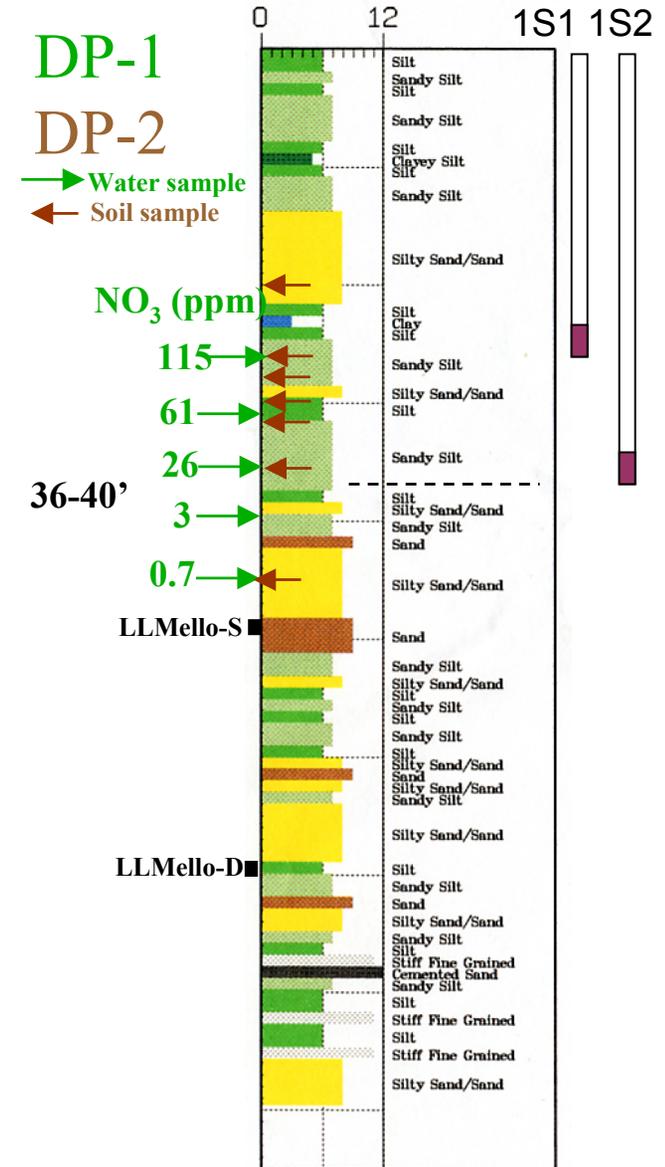
- Water samples: SS bailer (~500 mL)
  - 3 VOA vials for MIMS
  - 1 filtered 5 ml for IC
  - 125-mL or 1-L plastic for nitrate isotopics
- Soil samples: 12-inch sections
  - RNA: Dry ice/ethanol → dry ice (15-ml tube)
  - DNA: Dry ice or ice (plastic bag)

## Shallow water = irrigation?

- High nitrate; no excess N<sub>2</sub>; high DO
- High excess air; high dissolved CO<sub>2</sub>

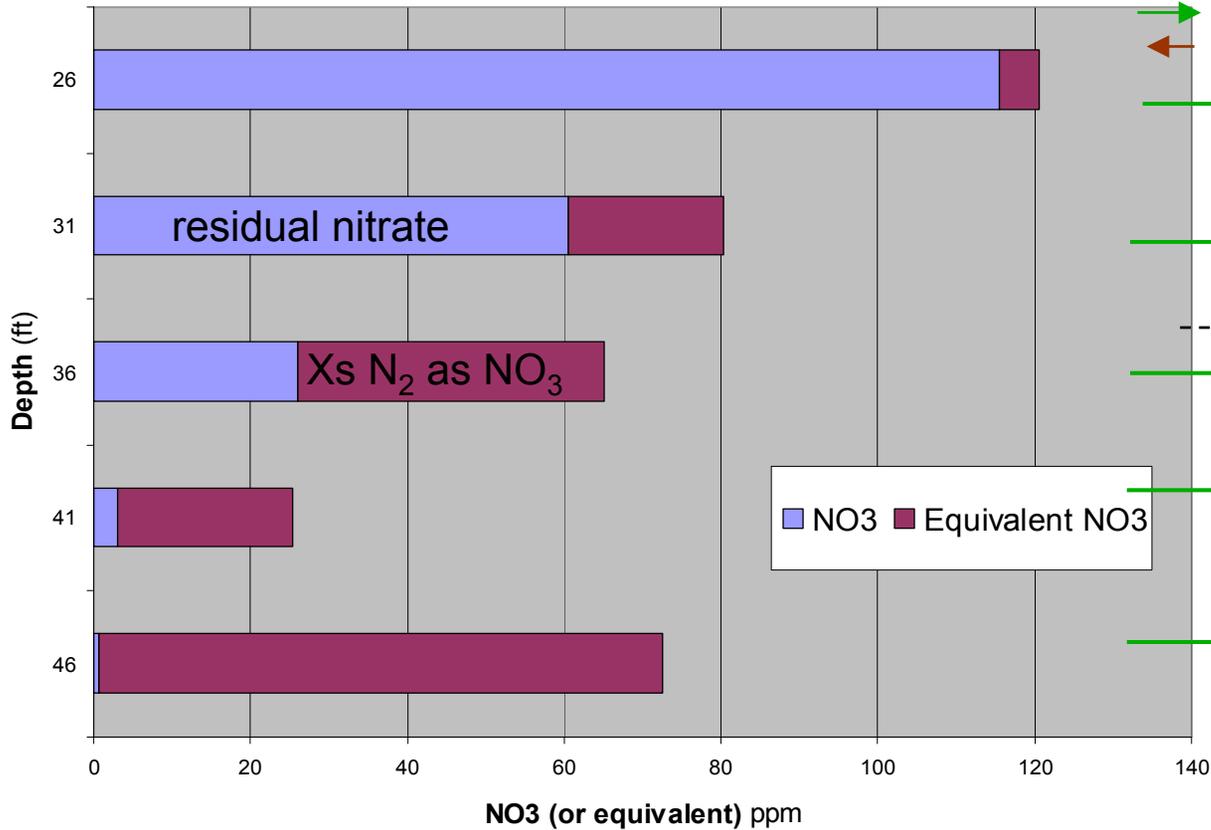
## Deeper water = mixed canal/irrigation?

- Low or no nitrate; variable excess N<sub>2</sub>; low DO
- Very low excess air; low dissolved CO<sub>2</sub>

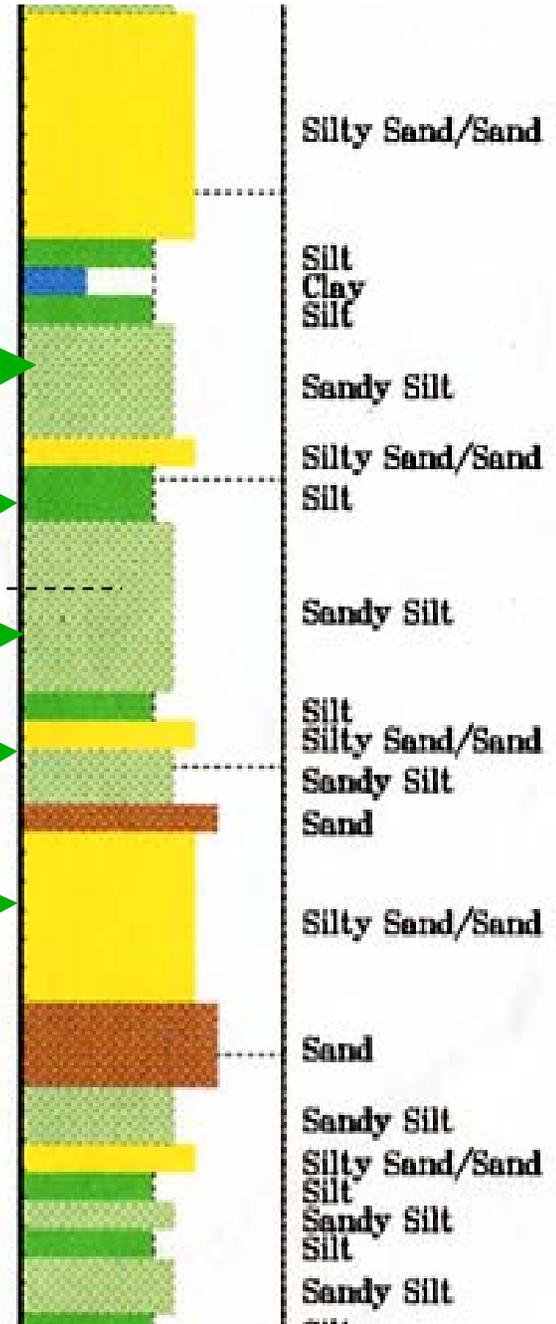


# Results of Excess Nitrogen Analysis:

DP1



Water sample



Silty Sand/Sand

Silt  
Clay  
Silt

Sandy Silt

Silty Sand/Sand  
Silt

Sandy Silt

Silt  
Silty Sand/Sand  
Sandy Silt

Sand

Silty Sand/Sand

Sand

Sandy Silt

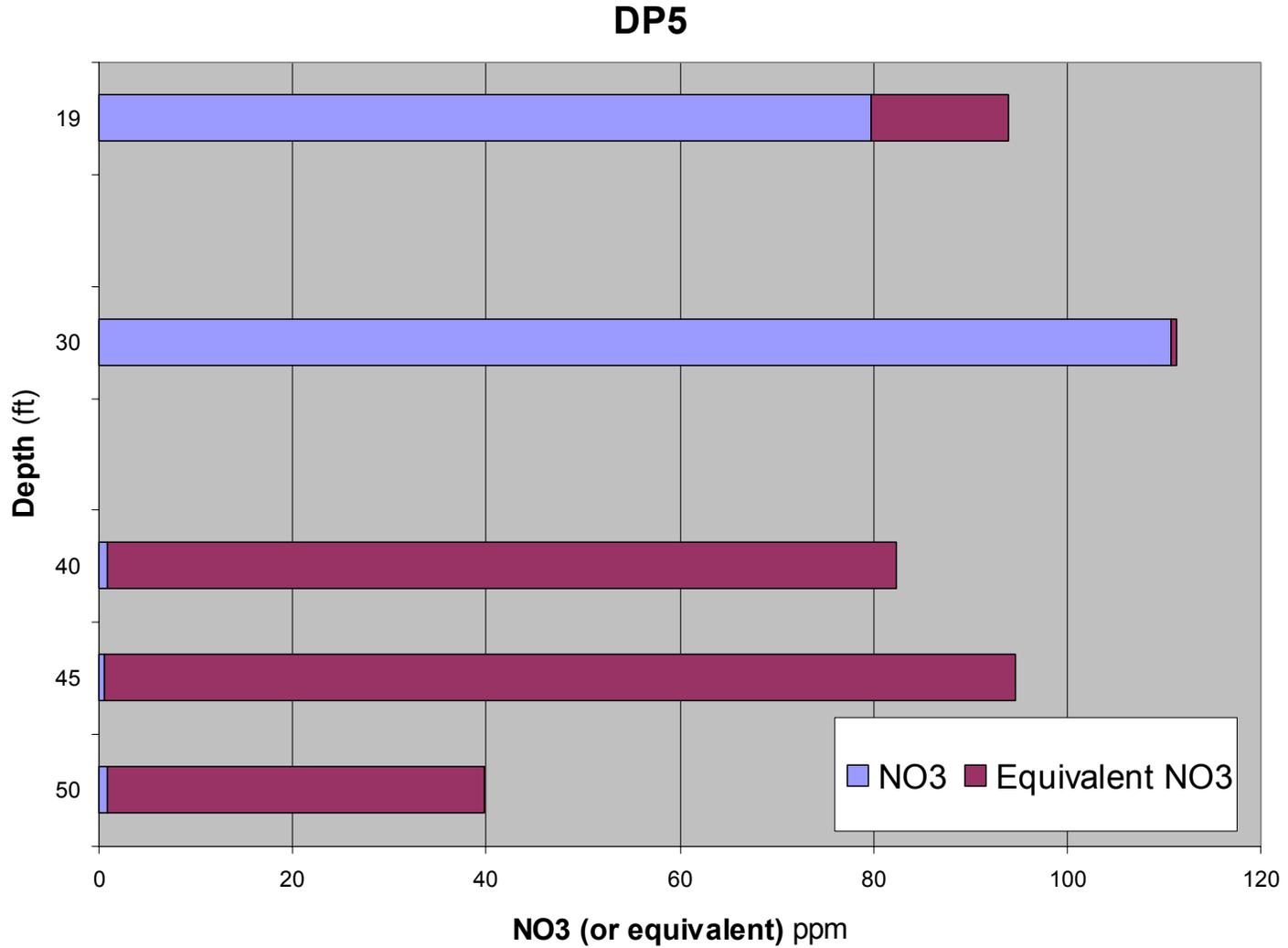
Silty Sand/Sand  
Silt

Sandy Silt  
Silt

Sandy Silt

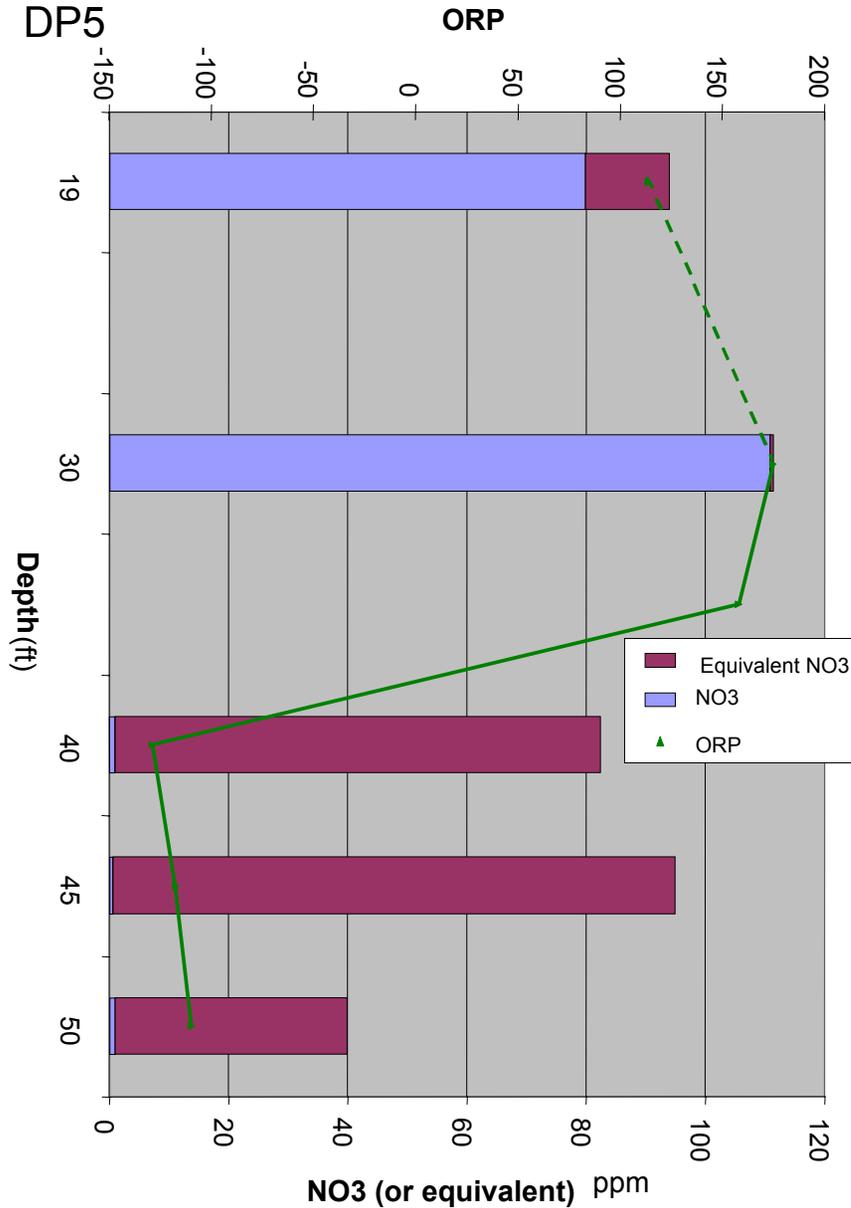
Silt

The same pattern occurs all across the dairy site.

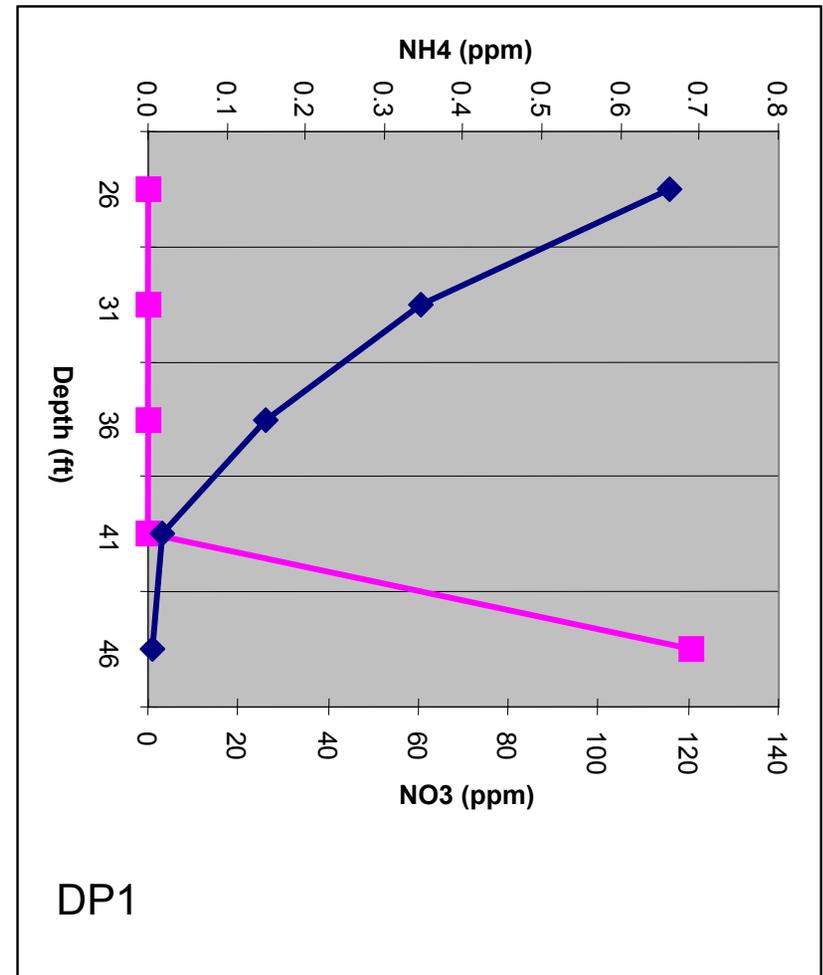




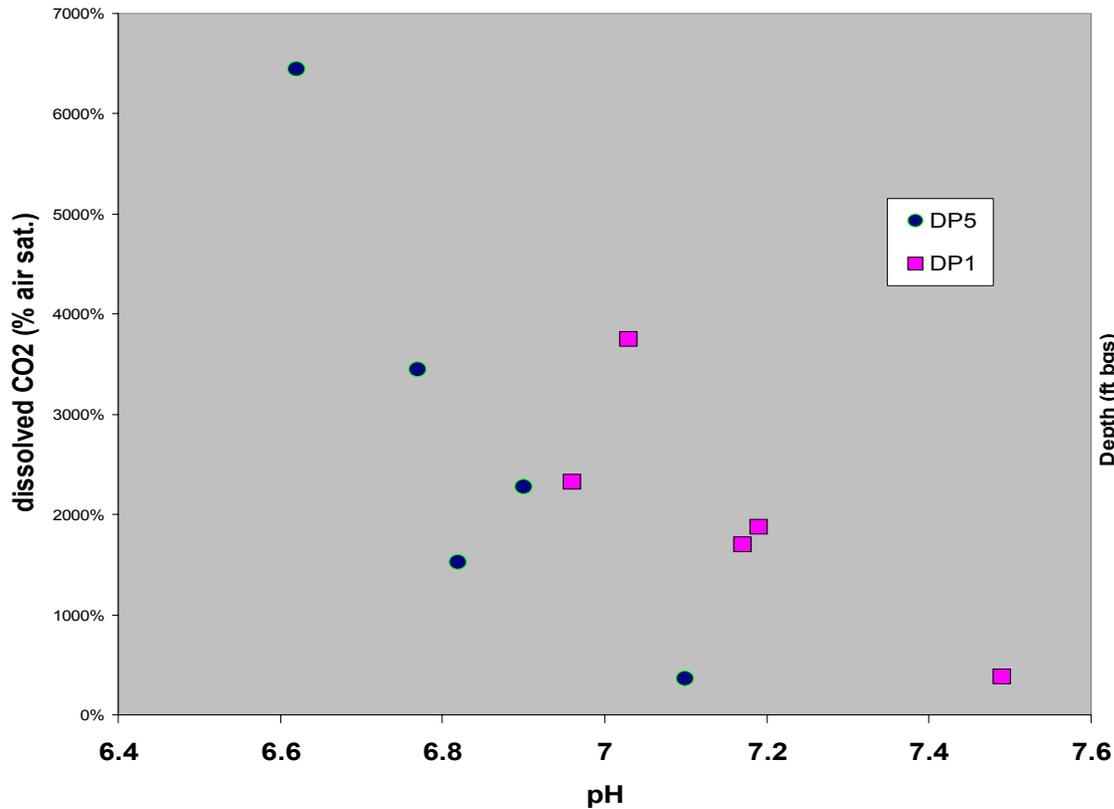
# We measured ORP, DO, pH, and other constituents



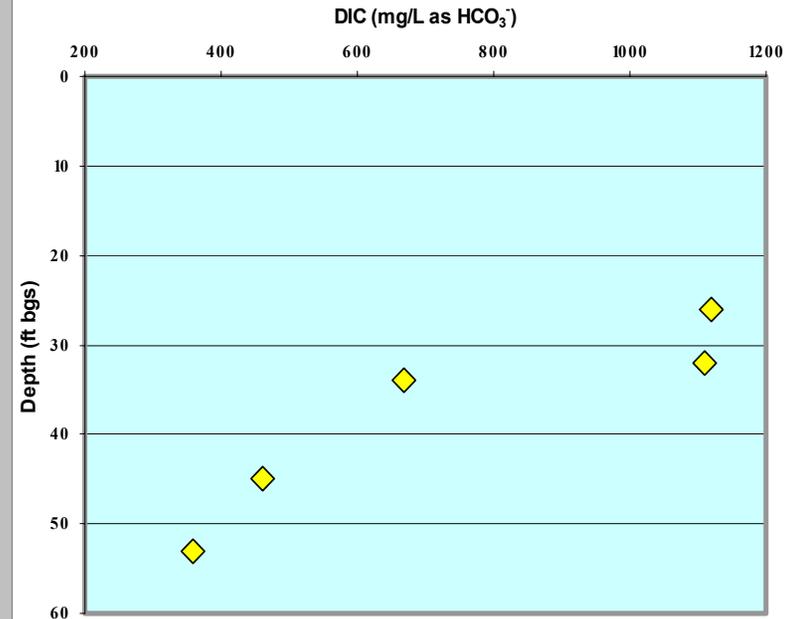
Field-measured ORP, nitrite, ammonium, and pH follow predictable trends



# Analytical results provide input for geochemical models

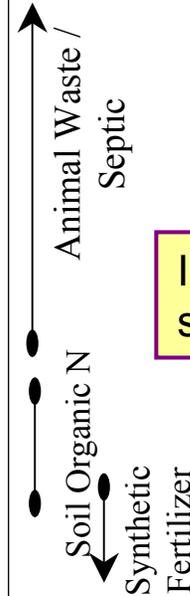
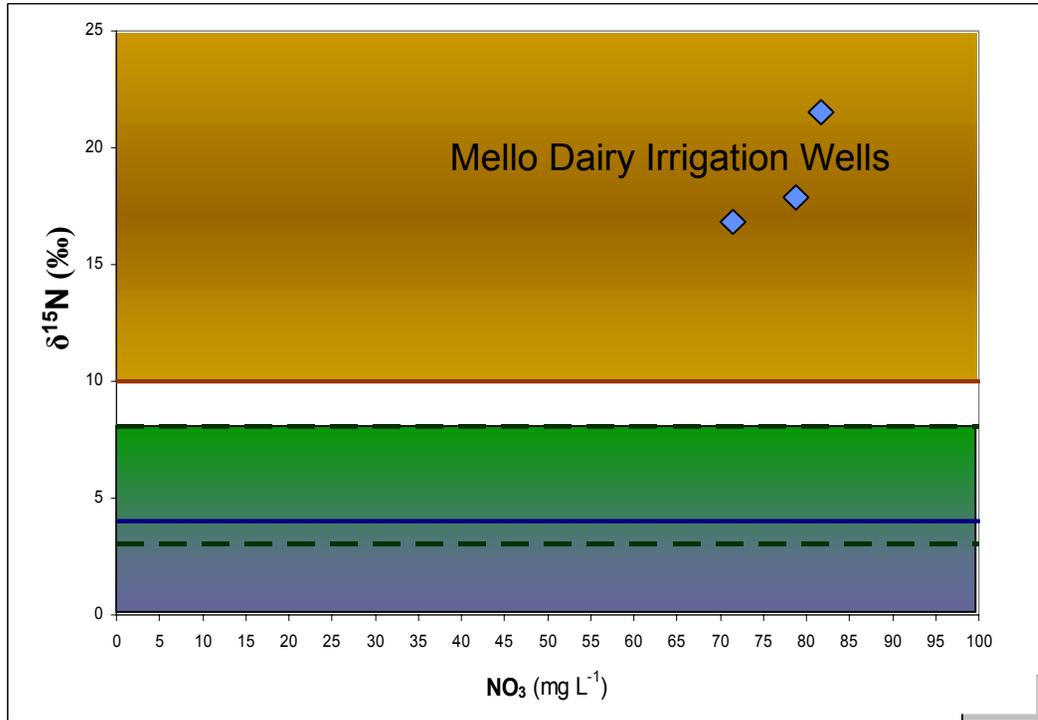


**Dissolved CO<sub>2</sub> is negatively correlated with pH**



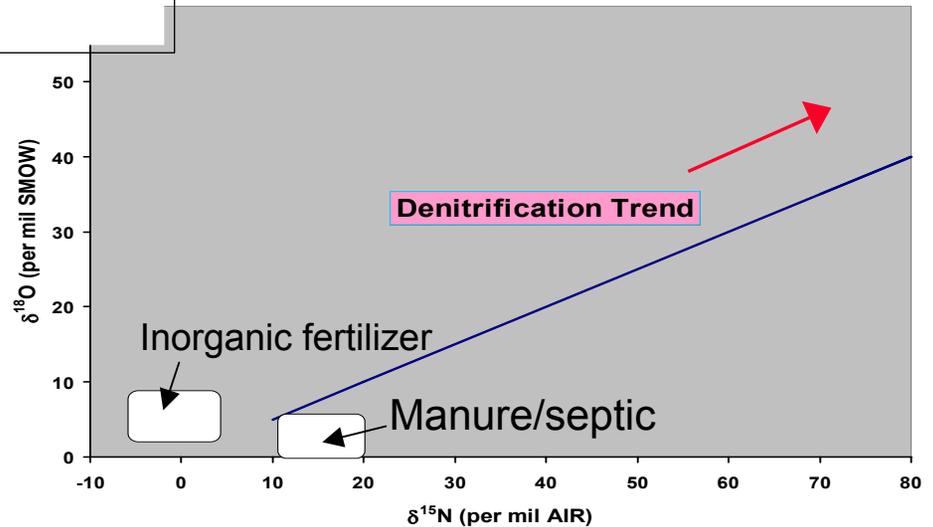
**Dissolved inorganic carbon decreases with depth**

# Nitrogen and Oxygen Isotopes of Nitrate indicate nitrate source and denitrification



Isotope signatures of nitrate sources may overlap

Changes in isotopic composition along a flowpath can indicate denitrification



# Using nitrate co-contaminants as tracers



Herbicides, pesticides, and fecal sterols are likely co-contaminants

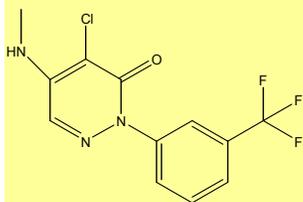
## Method Development

- Solid Phase Extraction suitable for selected target analytes
- GC-MS
- GC-ECD Dual column
- LC-MS/MS

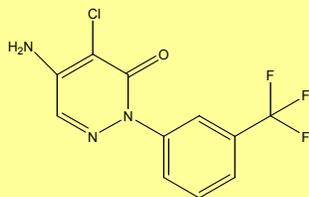
## Field Studies

- Lagoon source characterization for fecal sterols
- Identified triazine herbicides
- Identified norflurazon and desmethylnorflurazon

Herbicide-Chlorophyll/Carotenoid Pigment Inhibitor

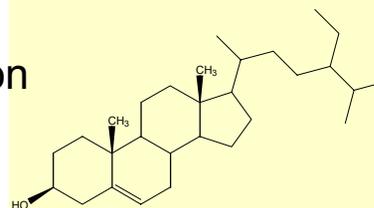


Norflurazon [27314-13-2]

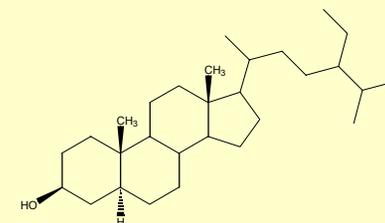


Desmethyl norflurazon [112748-69-3]

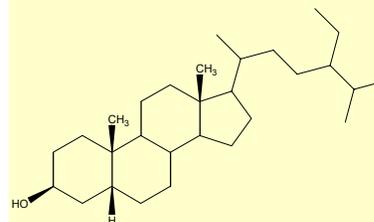
C<sub>29</sub> Sterols



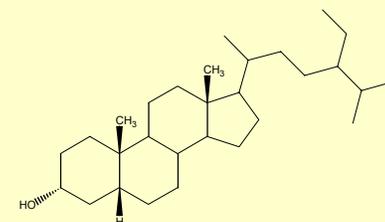
24-Ethylcholest-5-en-3β-ol (24-Ethylcholesterol or β-Sitosterol)



24α-Ethyl-5α-cholestan-3β-ol (24-Ethylcholestanol or Stigmastanol)



24-Ethyl-5β-cholestan-3β-ol (24-Ethylcoprostanol)



24-Ethyl-5β-cholestan-3α-ol (24-Ethyl-epicoprostanol)

# Conclusions



- **Excess nitrogen is a fast, accurate method for identifying and quantifying denitrification in groundwater**
- **Denitrification should be considered in CAFO regulations**
- **15 new shallow monitoring wells installed in September 2004**
  - **More accurate geochemical and isotopic data**
  - **Mixing of irrigation and canal water (age dating)**
- **Development of flow and transport models**
  - **Canal and irrigation recharge to perched aquifer**
  - **Use of CPT data as conditioning**
  - **Integration of kinetic models**
  - **Calibration to groundwater ages and measurements of DO, ORP, CO<sub>2</sub>, excess air and N<sub>2</sub>**
  - **Reactive transport modeling using analytical results**

